

# A new Spectrophotometric Method for Determination of Moxifloxacin with 1-(2-hydroxyphenylazo)-2,7-dihydroxynaphthalene in Bulk and Pharmaceutical Formulations

Rana M. W. Kazan<sup>1,\*</sup> Hassan Seddik<sup>2</sup>, Mahmoud Aboudane<sup>3</sup>

<sup>1</sup>Postgraduate student (PhD), Faculty of Science, Aleppo University, Syria

<sup>2</sup>Department of Chemistry, Faculty of Science, Aleppo University, Syria

<sup>3</sup>Department of Chemistry, Faculty of Science, Aleppo University, Syria

**\* Corresponding Author:**

Rana M. W. Kazan, Faculty of Science, Aleppo University, Syria. E-mail: Kazan \_ Rana @yahoo.com

**ABSTRACT:**

Moxifloxacin hydrochloride (MXF) is a fourth generation fluoroquinolone broad spectrum antibiotic agent used in conjunctivitis. A new, simple, rapid, and sensitive spectrophotometric method was developed for the determination of a (MXF). This method is based on the formation of ion-pair complex between the basic drug (MXF), and new dye; 1-(2-hydroxyphenylazo)-2,7-dihydroxy naphthalene (HPN). The formed complex was measured at 530 nm by using chloroform as solvent. The analytical parameters and their effects are investigated. Beer's law was obeyed in the range of 0.161 – 12.043 µg/mL, with correlation coefficient  $R^2 = 0.9998$ . The average recovery of Moxifloxacin was between 98.14 and 101.09 %. The limit of detection was 11.60 ng/mL and limit of quantification was 35.16 ng/mL. The proposed method has been successfully applied to the analysis of the studied drugs in pure forms and pharmaceutical formulations.

**Keywords:** Moxifloxacin hydrochloride; Spectrophotometer; pharmaceutical formulations.

## 1. INTRODUCTION

Moxifloxacin, 1-cyclopropyl-7-[(S,S)-2,8-diazabicyclo[4.3.0]non-8-yl]-6-fluoro-8-methoxy-1,4-dihydro-4-oxo-3-quinoline carboxylic acid is a synthetic bactericidal 4<sup>th</sup> generation fluoroquinolone (fluorinated derivative of the quinolone) discovered in 1996 and reached the UK market in 2000 [1-3].

Moxifloxacin is a broad spectrum important antibacterial agent used in human medicine which is active against broad spectrum of pathogens, encompassing Gram-negative, Gram-positive bacteria including resistant strain *Streptococcus pneumoniae*. It is also active against other microorganisms such as *Chlamydia pneumoniae* and *Mycoplasma pneumoniae* [4-7].

Various analytical techniques have been applied for the determination of MXF including: High performance thin layer chromatography HPTLC [8-10], Liquid chromatography mass spectrometry (LC/MS) [11], High performance liquid chromatography [HPLC-UV] [12-14], HPLC-mass spectrometry [15], Reverse phase high performance liquid chromatography [RP-HPLC] [16-19], Capillary electrophoresis [20], potentiometry [21,22], conductometry [23], voltammetry [24], polarography analytical [25], protonation equilibrium [26], Spectrofluorimetry [27,28], and spectrophotometric methods [29-34]. In the present work, we report the development of accurate and sensitive spectrophotometric methods based on the chloroform soluble ion-pair complexes between MXF and a new dye [35] 1-(2-hydroxyphenylazo)-2,7-dihydroxynaphthalene HPN (Fig.1). The absorbance measurements were measured at optimum wavelengths. The proposed method was

successfully applied for the determination of the studied drugs in pure and pharmaceutical formulations. No interference was observed from the additives. This method is morerapid, economic and sensitive in compared with the previously reported spectrophotometric methods, and it was validated by the statistical data.

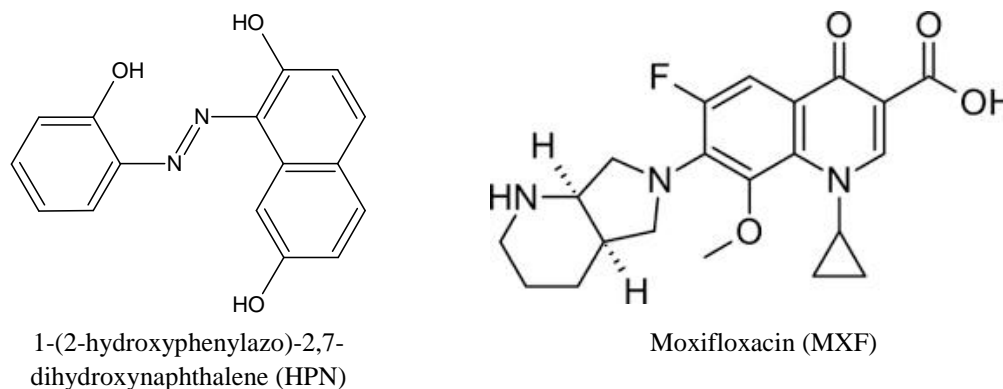


Fig. 1: The chemical structure of the studied compounds.

## 2. MATERIALS AND METHODS

### 2.1 Apparatus

UV/VIS double beam spectrophotometer V-630 (JASCO, Japan) equipped with 1 cm quartz cells; ultrasonic bath (Daihan, Korea); analytical balance TE64 (Sartorius, Germany) with accuracy  $\pm 0.1$  mg; digital pipettes (Accumax); drying oven (WTB binder-78532 TUTTLINGEN, Germany).

### 2.2 Reagents and Materials

All reagents and chemicals used were of analytical reagent or pharmaceutical grade.

Methanol and chloroform (99.0%, Sigma-Aldrich).

HPN ( $M_w = 280.00 \text{ g}\cdot\text{mol}^{-1}$ ) [35].

Moxifloxacin hydrochloride (MXF) (99.2%,  $M_w = 437.89 \text{ g}\cdot\text{mol}^{-1}$ , ROCHE PHARMA AG, Germany).

Syrian commercial dosage forms of MXF:

- ) Moxicin tablet (Ibn Al haytham) 400 mg of MXF.
- ) Moxiflox tablet (Razi) 400 mg of MXF.
- ) Moxaquin tablet (Obari) 400 mg of MXF.
- ) Megamox eye drope (Rama pharma), 5 mg of MXF/mL.

### 2.3 Preparation of solutions.

#### 2.3.1 . Preparation of Standard Stock Solution MXF ( $2.10^{-3} \text{ M}$ )

Standard Stock solution of Moxifloxacin was prepared by dissolving an accurately weighted 88.3 mg of MXF.HCL (including the purity, 99.2 %) in 100 mL of Methanol to get a suitable concentration. This solution was prepared fresh daily.

#### 2.3.2 Preparation of Standard Stock Solution (HPN) ( $2.10^{-3} \text{ M}$ )

Standard Stock solutions of (HPN) was prepared by dissolving an accurately weighed 56.0 mg in 100 mL of Methanol to get a suitable concentration. This solution are stable for at least one week if kept in the refrigerator.

#### 2.3.3 . Preparation of calibration curve

To prepare calibration standards, compatible volumes of stock standard solutions were diluted with chloroform into volumetric flasks (10 mL) to obtain final drug concentration of (0.161, 0.241, 0.321, 0.401, 0.482, 0.562, 0.642, 0.723, 0.803, 2.007, 4.014, 6.021, 8.029, 10.036, 10.839, 12.043)  $\mu\text{g/mL}$  and linearity was

studied. Linearity relationship was observed in the range 0.161 to 12.043  $\mu\text{g} / \text{mL}$  (Fig. 2) against a reagent blank as reference at 530 nm (Table 1).

### 3. Estimation of Moxifloxacin in commercial formulations.

#### 3.1 Estimation of Moxifloxacin in tablets.

For analysis of commercial formulations, ten tablets were weighed, powdered and then mixed well. Tablet powder equivalent to 400 mg of Formulation were transferred into 100 ml volumetric flask and dissolved in methanol. Then the solution was sonicated using ultrasonic for 30 minutes and filtered. 1 ml of the filtrate was taken and further diluted with methanol to 100 ml (to form 40  $\mu\text{g}/\text{mL}$ ). Then 1.25 ml of the last solution was diluted with chloroform to 10 ml (to form 5  $\mu\text{g}/\text{mL}$ ).

The absorbance of the prepared solutions were measured at 530 nm for Moxifloxacin solutions against blank and the drug content was estimated. The results are shown in table (3).

#### 3.2 . Estimation of Moxifloxacin in eye drops.

0.1 ml was taken from eye drop of the mixed contents of five eye drops and diluted with methanol to 10 ml (to form 50  $\mu\text{g}/\text{mL}$ ). Then 1 ml of the last solution was diluted with chloroform to 10 ml (to form 5  $\mu\text{g}/\text{mL}$ ). Table (3).

## 4. RESULTS AND DISCUSSIONS

### 4.1 Spectrophotometric analysis of Moxifloxacin with HPN.

The method is based on the formation of an orange ion-pair complex in chloroform, between the fluorquinolone MXF and the reagent HPN. The absorption spectra of the ion-pair complex, MXF-HPN, was measured in the range of 480– 600 nm against the blank solution. The complex has a maximum absorbance at 530 nm as shown in (Fig. 2).

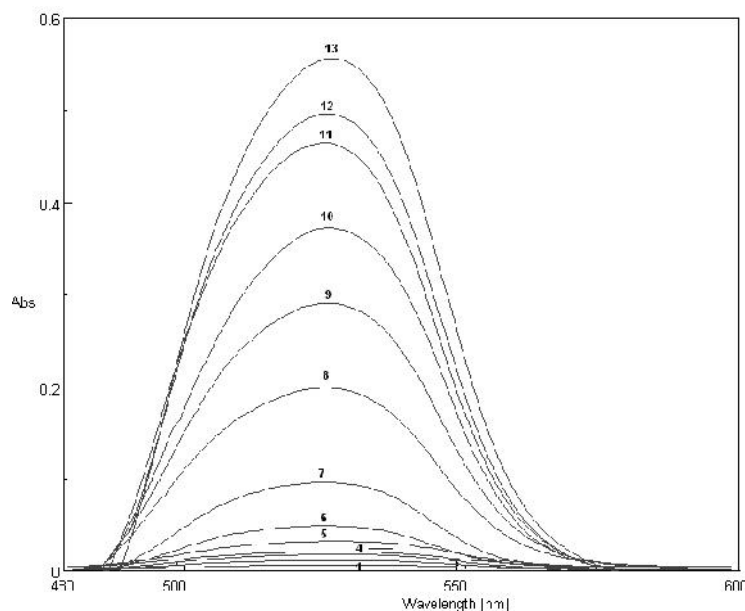


Fig.2: Spectra of complex MXF- HPN for various concentrations of MXF ( $\mu\text{g}/\text{mL}$ ) 1) 0.161, 2) 0.241, 3) 0.321, 4) 0.401, 5) 0.723, 6) 0.803, 7) 2.007, 8) 4.014, 9) 6.021, 10) 8.029, 11) 10.036, 12) 10.839, 13) 12.043

#### 4.2 Optimization of the reaction conditions.

The optimization conditions of the method were carefully studied to achieve complete reaction formation, highest sensitivity, and maximum absorbance.

##### 4.2.1 Effect of solvents.

The effect of several solvents, namely, chloroform, acetone, acetonitrile, benzene, diethyl ether, dichloromethane, dichloroethane, and tetrachloromethane, was studied in order to choose the effective solvent for complex which it had a maximum absorbance.

Chloroform was found to be the most suitable solvent for colored ion-pair complex. Experimental results indicated that the complex with total volume 10 mL chloroform, yielding maximum absorbance intensity, stable absorbance for the studied drugs and considerably lower extraction ability for the reagent blank. The optimization of the method was carefully studied to achieve complete reaction formation, high sensitivity, and maximum absorbance.

##### 4.2.2 Effect of Reagent Concentration.

The effect of the reagent was studied by measuring the absorbance of solutions containing a fixed concentration of MXF, varied amounts of HPN reagent. Maximum color intensity of the complex was achieved with 1.0 mL of  $2.0 \times 10^{-3}$  M of HPN solution, (Fig. 3).

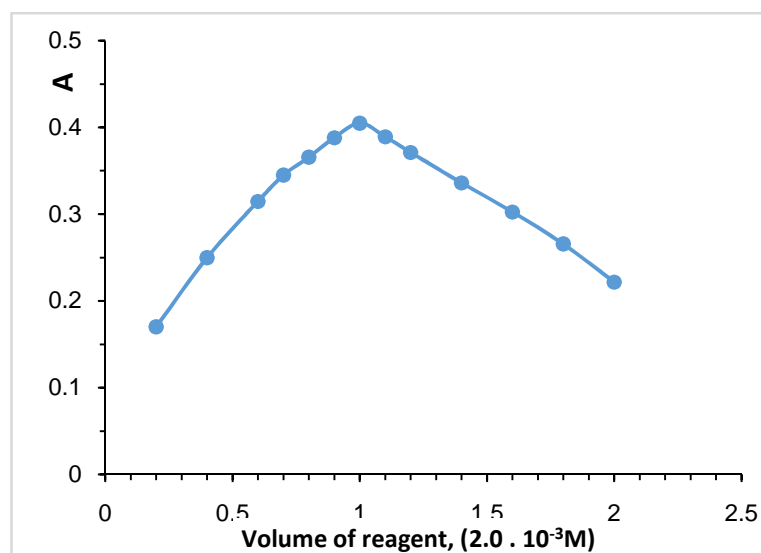


Fig.3: Effect of volume of ( $2.0 \times 10^{-3}$  M) reagent on the ion-pair complex MXF- HPN

##### 4.2.3 Effect of Time and Temperature.

The optimum reaction time was investigated from 0.5 to 5.0 min by following the colored development at ambient temperature ( $25 \pm 2^\circ\text{C}$ ). Complete color intensity was attained after 2.0 min of mixing of the complex. The effect of temperature on colored complexes was investigated by measuring the absorbance values at different temperatures. It was found that the colored complexes were stable up to  $35^\circ\text{C}$ . At higher temperatures, the drug concentration was found to increase due to the volatile nature of the chloroform. The absorbance remains stable for at least 10 h at room temperature.

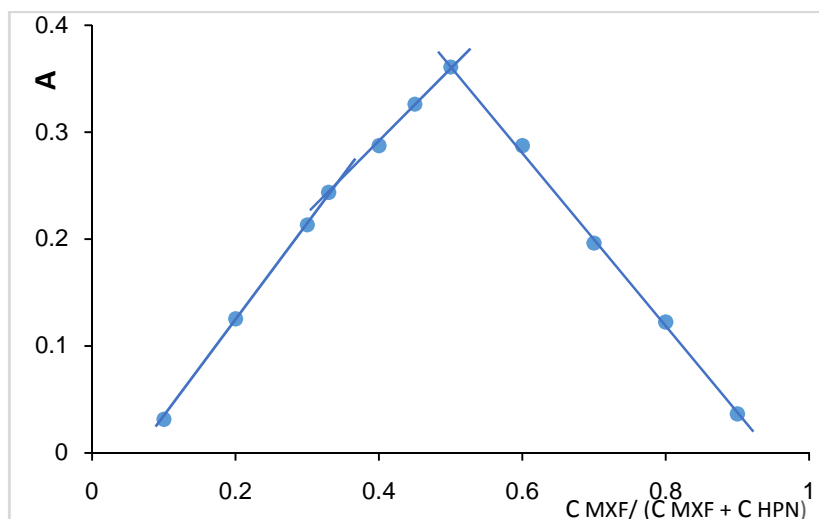
#### 4.3 Stoichiometric Relationship.

The stoichiometric ratio between drug and dye in the complex MXF-HPN was determined by Job's method of the continuous variation method, and Molar ratio method as following:

#### 4.3.1 Job's method of the continuous variation.

Job's method of continuous variation of equimolar solutions was employed:  $2.0 \times 10^{-4}$  M standard solution of MXF and  $2.0 \times 10^{-4}$  M solution of HPN was used. A series of solutions was prepared in which the total volume of MXF and HPN was kept at 2.0 mL and diluted with chloroform to 10 mL.

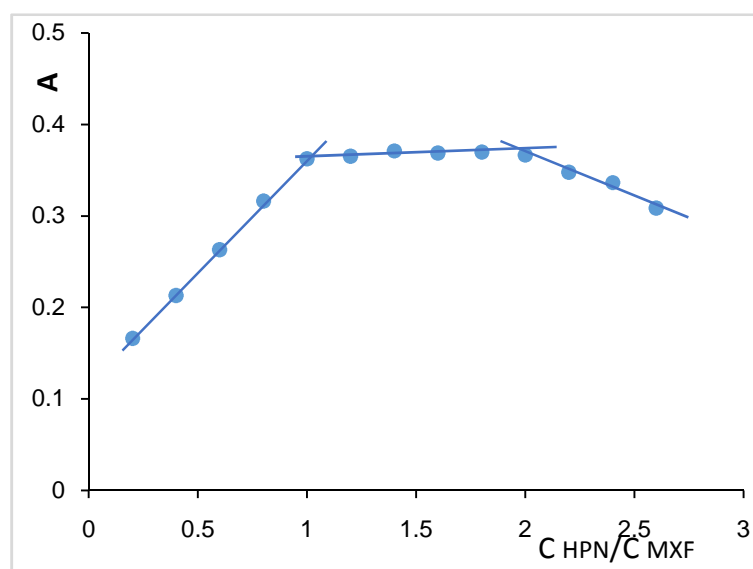
$A_{\max} = f([MXF]/[MXF]+[HPN])$ , The absorbance was measured at the optimum wavelength. The results showed two molar ratios 1:1 and 2:1 (HPN: MXF), (Fig. 4).



**Fig.4. Job's method of the continuous variation of MXF-HPN complex**

#### 4.3.2 Molar ratio method.

The stoichiometry of MXF-HPN complex by molar ratio method was studied according to the following equation:  $A_{\max} = f([HPN]/[MXF])$ , the results confirm the Stoichiometry of complex MXF-HPN that are 1:1 and 2:1 (HPN: MXF); Where the concentration of MXF is constant ( $2 \times 10^{-5}$  M), and the concentrations of HPN are changed from  $(0.4 \text{ to } 5.2) \times 10^{-5}$  M (Fig. 5).



**Fig.5. Molar ratio method of MXF-HPN complex**

## 5. METHOD VALIDATION

### 5.1 The linearity.

At described experimental conditions for MXF determination, standard calibration curves with reagent were constructed by plotting absorbance versus concentration. The absorbance obtained for the five analyses averaged at each concentration. The statistical parameters were given in the regression equation calculated from the calibration graphs. The linearity of calibration graphs was proved by the high values of the correlation coefficient ( $R^2 = 0.9998$ ) and the small values of the y-intercepts of the regression equations (fig.6). The apparent molar absorptivities of the resulting colored ion-pair complexes and relative standard deviation of response factors for each proposed spectrophotometric method were also calculated and recorded in Table1.

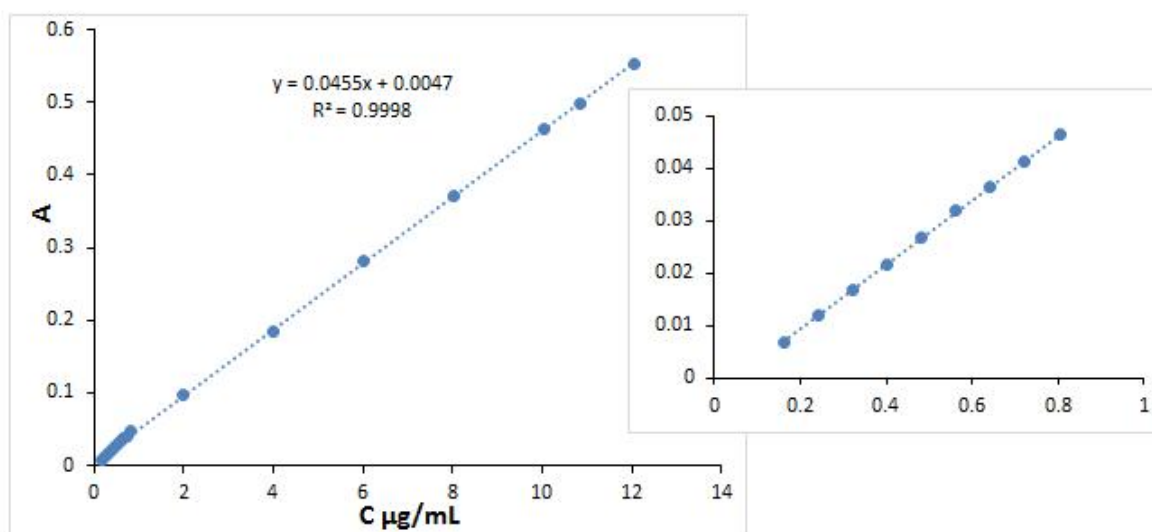


Fig. 6. Calibration curve for determination of MXF –HPN complex

Table 1: Spectrophotometric determination of MXF from MXF–HPN complex in chloroform

(µg/mL)		SD (µg/mL)	RSD%	Analytical standard error ASE X $\frac{SD}{\sqrt{n}}$ (µg/mL)	Confidence Limits $\bar{x} \pm t \frac{SD}{\sqrt{n}}$ (µg/mL)	Recovery %
X taken	X* found					
0.161	0.158	0.004	2.48	0.0018	0.158 ± 0.0050	98.14
0.241	0.239	0.005	2.07	0.0022	0.239 ± 0.0061	99.17
0.321	0.321	0.007	2.18	0.0031	0.321 ± 0.0086	99.95
0.401	0.400	0.008	1.99	0.0036	0.400 ± 0.0100	99.75
0.482	0.483	0.009	1.87	0.0040	0.483 ± 0.0111	100.00
0.562	0.567	0.011	1.96	0.0049	0.567 ± 0.0136	100.89

0.642	0.649	0.012	1.87	0.0045	0.649 ± 0.0150	101.09
0.723	0.727	0.013	1.80	0.0058	0.727 ± 0.0161	99.45
0.803	0.801	0.014	1.77	0.0063	0.801 ± 0.0175	99.75
2.007	2.013	0.024	1.19	0.011	2.013 ± 0.030	100.30
4.014	3.962	0.046	1.16	0.021	3.962 ± 0.058	98.70
6.021	6.070	0.062	1.02	0.028	6.070 ± 0.078	100.81
8.029	8.031	0.077	0.96	0.034	8.031 ± 0.094	100.02
10.036	10.055	0.086	0.85	0.038	10.055 ± 0.105	100.19
10.839	10.826	0.077	0.71	0.034	10.826 ± 0.094	99.88
12.043	12.022	0.082	0.68	0.037	12.022 ± 0.103	99.82

n = 5, t = 2.776

### 5.2 Sensitivity.

The limits of detection (LOD) and quantitation (LOQ) for the proposed method were found to be 11.60, 35.16ng/mL respectively, (Table 2).

**Table-2 The optimal spectrophotometric parameters for the determination of Moxifloxacin**

Parameter	Value
$\lambda_{\max}$ (nm)	530
Beer's law limits ( $\mu\text{g/mL}$ )	0.161 – 12.043
Molar absorptivity, (L/mol.cm)	20230.36
Regression equation	$Y = 0.0455X + 0.0047$
Slope (b)	0.0455
Intercept (C)	+ 0.0047
$R^2$	0.9998
LOD (ng/mL)	11.60
LOQ (ng/mL)	35.16
Sandell sensitivity SS ( $\mu\text{g/cm}^2$ )	0.0397

Note:  $Y = bX + c$ , where X is the concentration of drug in  $\mu\text{g/mL}$ .

### 5.3 Accuracy and Precision.

Percentage relative standard deviation (RSD%) and Analytical standard error (ASE%) of the suggested method were calculated and shown in Tables(1) These results of accuracy and precision show that the proposed method have good repeatability and reproducibility.

#### 5.4 Analysis of Pharmaceutical Formulations.

The proposed method has been successfully applied to the determination of MXF in pharmaceutical dosage forms in four trade mark Syrian products: Moxicin (Ibn Al haytham), Moxiflox (Razi), Moxaquin (Obari), and Megamox (Rama pharma). Table 3.

**Table-3**The obtained results for pharmaceuticals samples

Trade mark	Dose	$\bar{x}$ (mg/dose)	RSD%	Recovery %
Moxicin	400 mg/Tab	404.57	1.04	101.14
Moxiflox	400 mg/Tab	399.92	0.98	99.98
Moxaquin	400 mg/Tab	399.85	1.10	99.96
Megamox	5 mg/ml	5.04	1.53	100.76

Moreover, to check the validity of the proposed method, pharmaceutical dosage forms were tested for possible interference with standard addition method (Tables 4,5,6 and 7). There was no significant difference between slopes of calibration curves and standard addition methods. Therefore, it is concluded that the excipients in pharmaceutical dosage forms of MXF did not cause any interference in the analysis of MXF.

**Table-4**The recovery of Moxifloxacin in Moxicin

	*Sample µg/mL	*Added µg/mL	*Found µg/mL	RSD%	Recovery %
Moxicin	5	2	6.983	0.77	99.15
	5	4	8.991	0.74	99.77
	5	6	11.073	0.69	100.62

\* n = 5

**Table-5**The recovery of Moxifloxacin in Moxiflox

	*Sample µg/mL	*Added µg/mL	*Found µg/mL	RSD%	Recovery %
Moxiflox	5	2	7.021	0.80	101.05
	5	4	9.011	0.73	100.27
	5	6	11.052	0.65	100.87

\* n = 5

**Table-6** The recovery of Moxifloxacin in Moxaquin

Moxaquin	*Sample µg/mL	*Added µg/mL	*Found µg/mL	RSD%	Recovery %
----------	------------------	-----------------	-----------------	------	------------



	5	2	7.013	0.87	100.65
	5	4	8.986	0.82	99.65
	5	6	10.987	0.61	99.78

\* n = 5

**Table-7**The recovery of Moxifloxacin in Megamox

	*Sample µg/mL	*Added µg/mL	*Found µg/mL	RSD%	Recovery %
<b>Megamox</b>	5	2	7.033	0.77	100.40
	5	4	9.017	0.71	100.52
	5	6	11.047	0.65	100.23

\* n = 5

## 6. CONCLUSION

The new method is simple, rapid, economical, accurate and precise, and it is free from tedious experimental steps such as heating unlike the previously reported spectrophotometric methods cited earlier. The most attractive feature of this method is their relative freedom from interference by the usual diluents and excipients in amounts far in excess of their normal occurrence in pharmaceutical formulations. So it can be used for routine estimation of MXF from its pharmaceutical formulations.

## REFERENCES

- [1] S. Budavari, Eds, In; The Merck Index.13th edition, Merck and co., Inc., (White house station, NJ, 2001), 1097 and 1125.
- [2] M. Novitch (ed.), United States Pharmacopoeial Convention, Inc. (ed.), *The United States Pharmacopoeia (USP 34), The National Formulary (NF29)* (Elec. Ver., InSight Ver. 13.2 enigma, 2011).
- [3] M.I. Andersson, A.P. MacGowan, Development of the quinolones, *Journal of Antimicrobial Chemotherapy*, 51(Suppl. S1), 2003, 1–11.
- [4] P. Ball, *The quinolones* (history and overview. In: Andriole VT, ed. *The quinolones*, 3rd ed. San Diego: Academic Press, 2000, 1-31).
- [5] P.C. Appelbaum, and PA. Hunter, The fluoroquinoloneantibacterials: past, present and future perspectives, *Int. J. Anti microbial Agents*, 16, 2000, 5-15.
- [6] S. Sweetman, *Martindale: The Complete Drug Reference*, 37<sup>th</sup>Edn. (Pharmaceutical press, 4200pp, 2011).
- [7] E.C. Johannsen, M.S. Sabatine, *PharmCards: Review Cards for Medical Students*, Fourth Edition (Wolters Kluwer, Lippincott Williams & Wilkins, 2010).
- [8] V. Dhillon, and A. K. Chaudhary, A validated HPTLC method for estimation of Moxifloxacin hydrochloride in tablets, *African J of PharmaSci and Pharma*. 1(1),2010, 74-84.
- [9] S.K.Motwani, R.K. Khar, F.J. Ahmad, S. Chopra, K. Kohli, and S. Talegaonkar, Application of a validated stability-indicating densitometric thin-layer chromatographic method to stress degradation studies on Moxifloxacin, *Anal. Chim. Acta*, 582(1), 2007, 75-82.
- [10] M.V. Baldaniya, HPTLC method for estimation of Moxifloxacin in tablet dosage form, *Indian J.Pharma. Sciences*, 67(1), 2005, 112-115.
- [11] K. Vishanathan, M.G. Bartlett, and J.T. Stewart, Determination of Moxifloxacin in human plasma by liquid chromatography electro spray ionization tandem mass spectrometry, *J. Pharma.Biomed. Ana*, 30(4), 2002, 961-968.
- [12] J.A.O. González, M.C. Mochón, and F.J.B. De la Rosa, Simultaneous Determination of Cefepime and the Quinolones Garenoxacin, Moxifloxacin and Levofloxacin in Human Urine by HPLC-UV., *Mikrochim. Acta*, 151(1-2), 2005, 39-45.
- [13] L. Baietto, A. D'Avolio, F.G. De Rosa, S. Garazzino, S. Patanella, M. Siccardi, M. Sciandra, and G.D. Perri,

- Simultaneous quantification of Linezolid, Rifampicin, Levofloxacin and Moxifloxacin in human plasma using high-performance liquid chromatography with UV., *Ther. Drug Monit.*, 31(1), 2009, 104-109.
- [14] F.U. Khan, F. Nasir, Z. Iqbal, I. Khan, N. Shahbaz, M. Hassan, and F. Ullah, Simultaneous determination of Moxifloxacin and Ofloxacin in physiological fluids using high performance liquid chromatography with ultraviolet detection, *J Chromatogr B Analyt Technol Biomed Life Sci.* 2016, Mar 4.
- [15] B. Raju, M. Ramesh, R. M. Borkar, R. Padiya, S.K. Banerjee, R. Srinivas, Development and validation of liquid chromatography mass spectrometric method for simultaneous determination of moxifloxacin and ketorolac in rat plasma: application to pharmacokinetic study, *Biomed. Chromatogr.* 26(11), 2012, 1341-1347.
- [16] K. Kalyani, P. Sowjanya, T. Rajesh, and G.V. Kumar, Analytical Method Development and Validation by RP-HPLC for Simultaneous Estimation of Moxifloxacin and Prednisolone in Bulk Drug and Pharmaceutical Dosage, *Int. J. Chem. Pharm. Sci.*, 3(10), 2015, 2073-2080
- [17] A.P. Dewani, B.B. Barik, S.K. Kanungo, B.R. Wattyani and A.V. Chandewar, Development and Validation of RP-HPLC Method for the Determination of Moxifloxacin in Presence of Its Degradation Products, *American-Eurasian Journal of Scientific Research* 6 (4), 2011, 192-200.
- [18] P. Djurdjevic, A. Ciric, A. Djurdjevic, and M.J. Stankov, Optimization of separation and determination of Moxifloxacin and its related substances by RP-HPLC, *J. Pharm. Biomed. Anal.*, 50(2), 2009, 117-126.
- [19] P.R. Subbaiah, M.V. Kumudhavalli, C. Saravanan, M. Kumar, R.M. Chandira, Method development and validation for estimation of moxifloxacin HCl in tablet dosage form by RP-HPLC method, *Pharm. Anal. Acta*, v. 1, issue 2, 2010, 109.
- [20] A.F. Faria, M.V.N. de Souza, M.A.L. de Oliveira, Validation of a capillary zone electrophoresis method for the determination of ciprofloxacin, gatifloxacin, moxifloxacin and ofloxacin in pharmaceutical formulations, *J. Braz. Chem. Soc.*, 19(3), 2008.
- [21] A.A. Sakur, M.S. Bassmajei, and H.A. Dabbeet, Novel Moxifloxacin Ion Selective Electrodes for Potentiometric Determination of Moxifloxacin in Pure Form and Pharmaceutical Formulations, *International Journal of Academic Scientific Research* 3,(4), 2015, 66-75.
- [22] M.M. Hefnawy, A.M. Homoda, M.A. Abounassif, A.M. Alanazi, A.-r. Al-Majed, G.A. Mostafa, Potentiometric determination of moxifloxacin in some pharmaceutical formulation using PVC membrane sensors, *Chemistry Central Journal*, 8(59), 2014, 59-67.
- [23] S.M. Al-Ghannam, Atomic absorption spectroscopic, conductometric and colorimetric methods for determination of some fluoroquinolone antibacterials using ammonium reineckate. *Spectrochimica Acta Part A*, 69, 2008, 1188-1194.
- [24] M.A.G. Trindade, G.M. Da Silva and V.S. Ferreira, Determination of Moxifloxacin in tablets and human urine by square-wave adsorptive voltammetry, *Microchemical Journal*, 81, 2005, 209-216.
- [25] I. Recai, M. Hulya, Y. Emine and U. Bengi, Differential Pulse Polarographic Determination of Moxifloxacin Hydrochloride in Pharmaceuticals and Biological Fluids, *Anal. Lett.*, 40, 2007, 529.
- [26] M.H. Langlois, M. Montagut, J.P. Dubost, J. Grellet and M.C. Saux, Protonation equilibrium and lipophilicity of Moxifloxacin, *J. Pharm. Biomed. Anal.*, 37, 2005, 389-93.
- [27] J.A. Ocaña, F.J. Barragán, and M. Callejón, Spectrofluorimetric determination of Moxifloxacin in tablets, human urine and serum, *Analyst*, 125(12), 2000, 2322-2325.
- [28] U. H. Shah, Development and Validation of Spectrofluorimetric Method For The Estimation of Moxifloxacin in Pharmaceutical Dosage Form. *International Journal of Pharmacy and Pharmaceutical Sciences* 5(4), 2013, 252-254.
- [29] S. K. Sahu, M. d. Afzal Azamb, D. Sahu, and M. Banarjee, Spectrophotometric Estimation of Moxifloxacin in Bulk and its Pharmaceutical Formulations, *Pharmacologyonline* 2, 2011, 491-502.
- [30] R. Kant, R. Bodla, R. Bhutani, and G. Kapoor, Difference Spectrophotometric Method For Simultaneous Estimation of Moxifloxacin And Cefixime Trihydrate in Bulk and Combined Dosage Form. *International Journal of pharmacy and pharmaceutical sciences*, 7(9), 2015, 316-321.
- [31] N. Pradhan, H. Rajkhowa, H. Giri, and B. Shrestha, Simultaneous Spectrophotometric Estimation of Moxifloxacin Hydrochloride and Doxorubicin Hydrochloride, *Int J Pharm Pharm Sci*, 7(11), 2015, 21-26.
- [32] K.A.S.M. Attia, M.W. I. Nassar, M.A. Hasan, A Comparative Study of Different Approaches for Stability- Indicating Determination of Moxifloxacin Hydrochloride in Presence of Its Acidic-Induced Degradation Product, *Human Journals Research Article* 6 (4), 2016, 255-278.
- [33] K.N. Tarkase, S.S. Admane, N.G. Sonkhede, S.R. Shejwal, Development and validation of UV-spectrophotometric methods for determination of moxifloxacin hcl in bulk and pharmaceutical formulations, *Der PharmaChemica*, 4(3), 2012, 1180-1185
- [34] R. M. W. Kazan, H. Seddik, and M. Aboudane, Development and Validation of Spectrophotometric Method for Determination of Moxifloxacin HCL in Bulk and Pharmaceutical Formulations, *International Journal of Academic Scientific Research* will be published in June 2017.
- [35] M. Aboudane, and R. Kassab, Synthesis of New Azo Dyes Derived from 2,7-Dihydroxynaphthalene, *International Journal of Academic Scientific Research* 3 (4), 2015, 143-149.